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Note

Identification of O³-monoacetylmorphine in illicit heroin using gas chromatography-electron-capture detection and mass spectrometry^{*}

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Illicit drugs that are produced clandestinely often contain trace impurities. The identification and quantitation of these impurities is of importance for forensic purposes, especially in providing evidence of conspiracy¹⁻⁶. The trace compounds in illicit heroin which are used for forensic purposes include codeine and morphine and their acetylated by-products, namely, acetylcodeine, O⁶-monoacetylmorphine (Ia) and O³-monoacetylmorphine (Ib). Previous methods used to quantitate these substances have utilized gas chromatography with flame ionization detection (FID)^{1,6}. The presence of the isomer Ib was not established by this procedure, however, due to its co-elution with major heroin by-products, as well as its low levels in heroin. In addition, other studies have not satisfactorily demonstrated the presence of Ib in illicit heroin⁷⁻¹¹. Most of the derivative Ia results from hydrolysis of heroin (Ie),



(1)

- (a) $R_1 = H$; $R_2 = COCH_3$
- (b) $R_1 = COCH_3$; $R_2 = H$
- (c) $R_1 = COC_3F_7$; $R_2 = COCH_3$
- (d) $R_1 = COCH_1$; $R_2 = COC_3F_7$
- (e) $R_1 = COCH_3$; $R_2 = COCH_3$

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whereas most Ib results from the incomplete esterification of morphine with acetic anhydride¹².

In order to allow for the accurate quantitation of trace-level impurities in illicit heroin, gas chromatography-electron-capture detection (GC-ECD) methodology has been developed recently for codeine, morphine and O⁶-monoacetyl-morphine¹³. During the utilization of this method, a number of unidentified peaks appeared in the gas chromatograms of all of the illicit heroin samples examined. One of these peaks resulted from the presence of Ib.

In the work described here the samples were derivatized with heptafluorobutyric anhydride (HFBA) followed by GC-ECD quantitation. Earlier studies have reported the excellent response of ECD towards perfluorinated compounds^{14,15}. Following the quantitative analysis, mass spectral characterization of the O^{6-} and O^{3-} monoacetylmorphine derivatives (Ic, Id) was accomplished, applying the approach of Anggard and Sedvall¹⁶ used for the ECD quantitation of catecholamine metabolites.

EXPERIMENTAL

Apparatus

The quantitative analysis was done on a Perkin-Elmer 990 gas chromatograph equipped with a 63 Ni ECD and interfaced with an Infotronics CRS 208 integrator. The gas chromatograph was fitted with a 6 ft. \times 4 mm I.D. glass column packed with 3% OV-17 on Gas-Chrom Q (100–120 mesh). The column, injection and detector temperatures were maintained at *ca.* 230, 275 and 300°, respectively. Nitrogen was used as carrier gas at a flow-rate of 100–120 ml/min.

GC-mass spectrometry (MS) studies were performed on a Finnigan 3100D GC-MS quadrupole instrument equipped with a jet separator. The GC-MS instrument was fitted with a 3 ft. \times 2 mm I.D. glass column packed with 3% OV-17 on Gas-Chrom Q (80–100 mesh). Other parameters: emission current, 0.5 mA; electron energy, 70 eV; preamplifier sensitivity, 10^{-7} A/V; electron multiplier, 2.8 kV. The column was programmed from 180° with a helium carrier gas flow-rate of 25 ml/min.

Solvents, chemicals and chromatographic materials

The solvents used in this study were distilled in glass, and all of the chemicals were of reagent grade quality. Chromatographic materials were obtained from the usual commercial sources. The standard of O³-monoacetylmorphine was obtained from the Division of Pharmaceutical Chemistry, Food and Drug Administration (Washington, D.C., U.S.A.) and prepared by the method of Welsh⁷. All of the other standards were supplied by the Special Testing and Research Laboratory, Drug Enforcement Administration (McLean, Va., U.S.A.).

Quantitative analysis

A quantity of 1–10 mg of heroin was weighed accurately in duplicate into the bottom of a 13-ml conical centrifuge tube. 1.0 ml of acetonitrile (containing 0.20 mg/ml of chlorpromazine hydrochloride internal standard) was added and the tube was swirled to dissolve the sample. Then 50 μ l of HFBA were added and the tube was agitated to dissolve the HFBA and swirled occasionally for 15 min. Finally, 2.0 ml of

light petroleum and 5.0 ml of saturated sodium bicarbonate (pH 8.7; prepared fresh daily) were added. The tube was shaken vigorously for 5-10 sec, vented carefully and then centrifuged to clarify the layers. About 2-3 μ l of the petroleum layer were injected into the gas chromatograph under the conditions described earlier. If necessary, further dilutions were made with petroleum.

A working standard curve of O³-monoacetylmorphine was established by treating varying amounts (10-200 μ g) of it with HFBA and proceeding with the sample analysis as above. All of the calculations were based on relative peak areas of the O³-monoacetylmorphine-HFB and chlorpromazine hydrochloride internal standard.

GC-MS analysis

The derivatization method was modified for GC-MS analysis. The sample (ca. 30 mg) was dissolved in 3.0 ml of acetonitrile in a 13-ml glass-stoppered conical centrifuge tube. About 50 μ l of HFBA were added and the solution was allowed to react at room temperature for 5 min. About 6 ml of light petroleum followed by 5 ml of saturated sodium bicarbonate solution were then added. The mixture was shaken for ca. 10 sec to extract the derivatized heroin impurities into the petroleum layer. The latter was then removed and evaporated to dryness. The sample was



MINUTES

Fig. 1. Typical gas chromatogram indicating the presence of the heptafluorobutyrates of codeine, morphine, O⁶-monoacetylmorphine and O³-monoacetylmorphine.



Fig. 2. Mass spectrum of O3-monoacetylmorphine-HFB (Id).

redissolved in a small volume of anhydrous diethyl ether and injected into the GC-MS instrument. The procedure was used on standard O^3 - and O^6 -monoacetylmorphine, as well as on several illicit heroin samples.

DISCUSSION

Quantitative analysis

About twenty illicit, "uncut", brown and white heroin samples were subjected to analysis (see Fig. 1 for a typical sample chromatogram). All of the samples contained Ib, varying in content from 0.1 to 2%. In most of the samples the contents of



Fig. 3. Mass spectrum of O6-monoacetylmorphine-HFB (Ic).

Ia and acetylcodeine varied from 3 to 15%, those of morphine and codeine varied from 0.01 to 0.5%. It was interesting that, even in illicit samples of very high purity, Ib could be detected and readily quantitated.

Most of the Ia in illicit heroin is due primarily to the ready cleavage of the O^3 -acetyl group in heroin during hydrochloride formation and solvent removal. On the other hand, the majority of Ib is due to incomplete acetylation of morphine during the heroin synthesis. It has been observed that the formation of Ib is largely dependent upon the length and temperature of the reaction when using acetic anhydride. The product of reaction of morphine and acetic anhydride at room temperature is primarily O^3 -monoacetylmorphine¹².

It is evident from Fig. 1 that an accurate quantitation of Ib can be dependent upon the amount of Ia in the sample. Since Ib elutes closely after Ia, large quantities of the latter can tail into the former, thus affecting the accuracy of the quantitation. Therefore, the chromatographic conditions may have to be modified in order to effect an accurate quantitation of Ib in some samples.



Fig. 4. Fragmentation of the O^3 -acetate and O^6 -heptafluorobutyrate esters of O^3 -monoacetyl-morphine-HFB (Id), showing the formation of the major ions.

The HFBA derivatives in light petroleum were stable for several hours. However, it is recommended that, upon derivative formation, the analysis be completed without delay.

Mass fragmentation of O^3 -monoacetylmorphine-HFB (Id) and O^6 -monoacetylmorphine-HFB (Ic)

The mass spectra of standard samples of Ib and Ia after derivatization with heptafluorobutyric anhydride were obtained (Figs. 2 and 3). The perfluorinated derivatives were then observed by GC-MS in illicit heroin. The presence of the perfluorinated group was supported by strong fragments at m/e 31 due to the [CF]⁺ fragment, m/e 69 due to the [CF₃]⁺ fragment and m/e 81 due to the [C₂F₃]⁺ fragment. Both Id and Ic produced readily recognizable molecular ions at m/e 523. However, fragmentation of the ester groups (acetate and heptafluorobutyrate) at different positions in the molecules produced different major ions which could in turn be used to distinguish the two isomers.

O³-Monoacetylmorphine–HFB produced a prominent ion at m/e 481 [M – (CH₂CO)][‡] from O³-acetate fragmentation, m/e 310 [M – (CF₃CF₂CF₂CO₂)][‡] from fragmentation of the O⁶-perfluorinated moiety and m/e 268 [M – (CH₂CO + CF₃CF₂CF₂CO₂)][‡], resulting from both fragmentation processes (Fig. 4). The O⁶-acetyl group of O⁶-monoacetylmorphine–HFB fragmented to give a base peak at m/e 464 [M – CH₃CO₂][‡] (Fig. 5). The heptafluorobutyryl portion at the O³ position fragmented to give m/e 326 [M – (CF₃CF₂CF₂CO)][‡], although only of minor intensity. Cleavage of the ethanamine bridge in the morphine moiety, [M – (CH₃CO₂ + CH₂=CH–NHCH₃)][‡], would produce the fragment at m/e 407 (ref. 17) (Fig. 5). Finally, the fragment at m/e 411 could reasonably have formed via the retro Diels–



Fig. 5. Fragmentation of the O⁶-acetate and ethanamine bridge of O⁶-monoacetylmorphine-HFB (Ic), showing the formation of the major ions.



Fig. 6. Formation of the ion at m/e 411 from O⁶-monoacetylmorphine-HFB (Ic) via a retro Diels-Alder mechanism.

Alder route shown by Audier *et al.*¹⁸ (Fig. 6). This involves loss of the acetoxyl group and ring C carbon atoms 6, 7, 8 and 14 ($-C_6H_8O_2$; 112 a.m.u.). The resulting fragment then retains the heptafluorobutyrate at the O³ position.

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